central noradrenergic and serotonergic activity. In animals it has appetite-stimulating and antinausea properties. These effects are attributed to the 5-HT₃ antagonist properties; therefore, it may share similar effects as antiserotonin medications discussed in Section Serotonin Antagonists.

Clinical Use

Mirtazapine has been studied more in cats than in dogs (Quimby and Lunn, 2013). These studies in cats demonstrated that it is an effective appetite stimulant. The dose is usually 1.88 mg per cat oral. At high doses it produces adverse effects that include vocalization and increased restlessness. In cats it has a half-life of approximately 10 hours, which allows for once-daily dosing. In cats with chronic kidney disease - for which appetite stimulation often is desired - clearance is slower and the half-life increases to 15 hours, which indicates that an every-other-day dosing schedule should be used in cats with kidney disease to avoid accumulation (Quimby et al., 2011). Mirtazapine is available as 7.5-mg tablets, but also is available in 15, 30, and 45-mg sizes. The formulation that some veterinarians prefer is a rapidly disintegrating oral tablet that dissolves easily in an animal's mouth (15, 30, and 45 mg).

Gastrointestinal Prokinetic Drugs

Prokinetic drugs increase gastrointestinal motility (Washabau and Hall, 1997). They are used in dogs, cats, horses, and occasionally ruminants to stimulate gastric emptying, rumen motility, or to increase intestinal motility (Whitehead et al., 2016). Intestinal motility is sometimes decreased after intestinal disease or surgery and can lead to ileus. Some of these drugs are intended to restore normal motility to facilitate recovery.

Metoclopramide (Reglan[®], Maxeran[®])

Metoclopramide has multiple actions. It is a dopamine (DA_2) antagonist, serotonin $(5-HT_4)$ agonist and serotonin $(5-HT_3)$ antagonist. Among the proposed mechanisms of metoclopramide is an increase in the release of acetylcholine in the GI tract, possibly via a prejunctional mechanism. It also may increase motility of gastric smooth muscle by increasing sensitivity of the cholinergic response. Since it also is a dopamine antagonist, it may antagonize dopamine's (DA_2) inhibitory action on GI motility.

Metoclopramide increases gastric emptying, increases the tone of the esophageal sphincter, and stimulates motility of the duodenum. It has less effect on distal segments of the intestine. Metoclopramide acts centrally to inhibit DA₂, which produces the antiemetic effects discussed in Section Antiemetic Drugs. In people, metoclopramide also has been used to treat hiccups and lactation deficiency.

Adverse effects from metoclopramide can include excitement (seen in horses, for example), anxiety, and involuntary muscle movements. There are also endocrine effects: There is a transient increase in prolactin and aldosterone. *Since some breast cancers are prolactin-dependent, there has been some concern about the carcinogenicity of this drug in women.*

Use in Small Animals

In dogs metoclopramide has been used as an antiemetic more commonly than other drugs. Although it has been used to promote GI motility as well, this effect is less established than previously thought (Whitehead et al., 2016). For example, it is of little benefit to increase stomach emptying in disorders of gastroparesis or chronic regurgitation. It also has been used to stimulate normal upper motility following surgery (e.g., corrective surgery for gastric dilatation), but one study showed that metoclopramide did not change gastric motor activity to promote gastric emptying in dogs with gastric dilatation volvulus (Hall et al., 1996). In another study, it reduced, but did not prevent gastroesophageal reflux in anesthetized dogs at a dose of 1 mg/kg (Wilson et al., 2006). Doses are in the range of 0.25 to 0.5 mg/kg, q 8-12 h, but they have been increased to 1-2 mg/kg.

Use in Horses

Some equine surgeons have used infusions of metoclopramide (0.125–0.25 mg/kg/h) added to IV fluids to reduce postoperative ileus in horses (Gerring and Hunt, 1986). It may stimulate small intestine – but not large bowel – motility, but this has little benefit for horses with intestinal ileus (Sojka et al., 1988). Undesirable side effects in horses have been common, and include behavioral changes and abdominal pain. Since this drug transiently increases prolactin secretion, there has been interest in using this drug for treating agalactia in animals, but efficacy has not been determined. Domperidone is preferred for this effect (see Section Domperidone (Motilium, Equidone)).

Use in Ruminants

The clinical use of metoclopramide in large animals has not been as common as in small animals. Metoclopramide has little usefulness in cattle, although it may increase the motility of the rumen in cattle and sheep. It has been used successfully in some cattle with functional pyloric stenosis (Braun et al., 1990), but was not effective in calves (0.1 mg/kg IM). At doses higher than 0.1 mg/kg in calves it caused severe neurological side effects (Wittek and Constable, 2005).

Domperidone (Motilium, Equidone)

Domperidone is a dopamine-2 receptor (DA_2) antagonist. It may also have α_1 -receptor antagonist and serotonin $(5-HT_2)$ antagonist effects. It has been available as a 10-mg tablet outside the USA as a human prokinetic drug but not allowed for human use in the USA because of cardiac toxicity. Its mechanism of action and GI prokinetic effects are similar to metoclopramide, but its efficacy has not been very impressive in animals and thus a clinical use has not been recommended (Whitehead et al., 2016). A difference between metoclopramide and domperidone is that the latter does not cross the blood-brain barrier. Therefore, adverse CNS effects are not as much of a problem compared to metoclopramide in horses. It may have antiemetic properties, but only if the stimulus for vomiting affects the CRTZ. It is capable of reaching the area postrema of the brain because this area is not protected by the blood-brain barrier. An additional effect is to stimulate lactation (see Section Use in Horses).

Use in Small Animals

The use is not reported, but it will produce a prokinetic effect in dogs at a dose of 0.05-0.1 mg/kg (2–5 mg/animal).

Use in Horses

Domperidone has been investigated for use in horses to treat fescue toxicity and agalactia. Fescue toxicosis is caused by a fungus that produces a toxin that induces reproductive toxicity in horses. The action of domperidone to increase lactation is through the stimulation of prolactin. It is approved by the FDA as an equine formulation of domperidone (Equidone oral gel, 11%). The approved dose is 1.1 mg/kg once daily starting 10–15 days prior to the anticipated foaling date. Treatment may be continued for up to 5 days after foaling if mares are not producing adequate milk. (This dose is equivalent to 5 ml per 500 kg – 5 ml per horse – daily, PO of the 11% oral gel.) Do not administer with stomach antacids such as omeprazole, cimetidine, or antacids.

The prokinetic effects in horses are not very impressive. At an IV dose of 0.2 mg/kg it was effective at restoring motility in horses with ileus, but this drug is not available in an injectable formulation. The oral absorption in horses is only 1.2–1.5%. Oral administration of 1.1 mg/kg (the approved dose) had no effect on GI function in horses but at 5 mg/kg it increased stomach emptying (Nieto et al., 2013).

Another use of domperidone is to increase digital laminar microvascular blood flow in horses. This effect is presumed to be via the action as an antagonist on vascular α_2 -adrenergic receptors. It was shown to increase laminar microvascular blood flow in normal horses (1.1 and 5.5 mg/kg oral), but has not been evaluated clinically for treatment of laminitis (Castro et al., 2010).

Cisapride

In July 2000, cisapride (formerly called Propulsid[®]) was removed from the market because of serious cardiac adverse events, and some deaths in people, secondary to cardiac arrhythmias. The drug sponsor has no plans to market this drug to veterinarians, but there is continued interest among veterinarians and it is still available via compounding pharmacists. Until other new replacement drugs become available, such as prucalopride or mosapride, veterinarians will rely on compounded formulations or consider alternative drugs.

The reviews on cisapride by Washabau and Hall (1995) and Van Nueten and Schuurkes (1992) describe the details of its mechanism of action and clinical effects. Cisapride has greater prokinetic effects in comparison to the other drugs discussed thus far. Its mechanism is believed to be as an *agonist* for the 5-hydroxytryptamine $(5-HT_4)$ receptor on myenteric neurons (5-HT₄ ordinarily stimulates cholinergic transmission in the myenteric neurons). (Serotonin and antagonists/agonists are covered in more detail in Chapter 19.) Cisapride may also be an antagonist for the 5-HT₃ receptor. Via these mechanisms or independently - cisapride may enhance release of acetylcholine at the myenteric plexus. There is evidence that, in cats, cisapride directly stimulates smooth muscle motility via an unknown noncholinergic mechanism (Washabau and Summarco, 1996). Cisapride increases the motility of the stomach, increases stomach emptying, and increases motility of the small intestine and colon. It accelerates the transit of contents in the bowel and intestines. Because of the 5-HT₃ antagonist properties, it also has some antiemetic effects. Other drugs with a similar mechanism of action have been investigated, but are not in clinical use. One such drug is mosapride. Like cisapride, mosapride is also a 5-HT₄ agonist and has been approved in some countries for treating upper gastrointestinal motility disorders in dogs (Chae et al., 2015). It has been studied in experimental horses and demonstrated to increase motility of the small intestine and cecum at a dose of 1.5-2 mg/kg PO (Sasaki et al., 2005).

Pharmacokinetics

Oral absorption is variable because of extensive metabolism. The oral absorption in dogs and cats ranges from 30 to 60%. In horses, rectal absorption has been attempted, but the amount absorbed systemically is negligible (Cook et al., 1997).

Elimination half-life is variable, but ranges from an average of approximately 5 hours in dogs and cats to a much faster rate in large animals, 2 hours or less in horses and ruminants. The volume of distribution is high in small animals (>4 l/kg) and approximately 1.5 l/kg in large animals.

Use in Dogs

In dogs at a dose of 0.1 mg/kg (range 0.08–1.25 mg/kg) orally, it stimulates smooth muscle of the stomach, small intestine, and colon, with a duration of effect of about 3 hours. Routine clinical doses have ranged from 0.1 to 0.5 mg/kg every 8–12 hours.

Although cisapride has been used by some veterinarians for treatment of megaesophagus in dogs, the response is usually poor. The canine esophagus is striated muscle, with no smooth muscle to directly respond to the medication. Clinical use in dogs has included treatment for gastroesophageal reflux, delayed gastric emptying, and small bowel motility disorders. Compared to metoclopramide, cisapride is more effective for increasing lower esophageal sphincter tone in dogs, which is helpful for preventing reflux esophagitis (Kempf et al., 2014).

Use in Cats

Experiments have demonstrated that cisapride causes stimulation of the entire GI tract in cats. Of particular interest is the effect of cisapride on colonic smooth muscle. Cisapride will stimulate this motility and has been used for treating chronic constipation. By contrast, metoclopramide has no effect on colonic smooth muscle. The dose of cisapride in cats is approximately 2.5 mg per cat, two or three times daily. Doses as high as 1 mg/kg every 8 hours, or 1.5 mg/kg every 12 hours have been recommended by some investigators (LeGrange et al., 1997).

Use in Horses

In horses cisapride increases the motility of the left dorsal colon and improves ileocecocolonic junction coordination. In contrast to metoclopramide, cisapride has fewer side effects at doses needed to affect the GI tract and greater effects on the jejunum and colon than metoclopramide. Many investigators believe that it has a place in the postoperative management of horses that have undergone abdominal surgery. One dose tested to be effective was 0.1 mg/kg, IV. At this dose, the effects appear to persist for approximately 2 hours. Oral administration is usually not possible in these horses because of gastric reflux and absorption after oral administration in a horse with gastric reflux probably is questionable.

Availability of Formulations

The previously available tablet was a 10-mg tablet from Janssen Pharmaceutica. Although cisapride is insoluble in most aqueous solutions, solubility is possible in acidic solutions. An IV form may be created by preparing a 4 mg/ml solution in tartaric acid by a reputable compounding pharmacist. The preparation of this formulation was described in the publication by Cook et al. (1997). To prepare this solution, 40 mg of cisapride is combined with 1 ml of 0.4 M tartaric acid. After the cisapride is dissolved, dilute with water to obtain a total volume of 10 ml. Oral formulations for cats have been prepared from the bulk powder administered in a capsule, via a suspension in a flavored vehicle or dissolved in cod liver oil.

Side Effects and Interactions

Adverse effects have not been reported in animals; however, abdominal discomfort has been observed when animals received high doses. In safety studies, dogs have tolerated high doses (40 mg/kg) for prolonged periods without problems.

In people, high plasma concentrations have caused cardiac arrhythmias. The arrhythmias are caused by prolonged QT intervals, presumably from blockade of potassium channels. This can lead to serious arrhythmias and has been responsible for deaths in people. These reactions have not been reported for animals. Nevertheless, one should be cautious about combining cisapride with drugs such as itraconazole and ketoconazole that may increase plasma concentrations by interfering with metabolism.

Bethanechol (Urecholine)

Many of the formulations of bethanechol have been discontinued and are no longer marketed. Some generic forms may still remain and veterinarians have also obtained it through compounding pharmacies. This drug is a cholinergic agonist that has been used to nonspecifically stimulate smooth muscle. It binds to muscarinic receptors and initiates GI smooth muscle contractions, but its actions are nonspecific. In contrast to cisapride or metoclopramide, bethanechol has a more pronounced effect on motility of the ileocecocolic region in cattle (0.7 mg/kg). In horses, bethanechol increases gastric emptying at a dose of 0.025 mg/kg IV (Ringger et al., 1996). One of its other uses has been to stimulate contraction of bladder smooth muscle in animals that have a failure to completely empty their urinary bladder when voiding. Adverse effects are common and include diarrhea and other consequences of cholinergic stimulation.

Neostigmine (Prostigmin)

Neostigmine inactivates the enzyme acetylcholinesterase, which results in inhibition of degradation of acetylcholine at the synapse. It prolongs the action of acetylcholine and may directly stimulate cholinergic receptors. It is short acting. In horses, its use is discouraged because it may actually decrease intestinal propulsive contractions, delay gastric emptying, and cause abdominal discomfort.

One of the other uses of neostigmine in animals is for the treatment of neuromuscular diseases such as myasthenia gravis. Its adverse effects are significant, and include diarrhea, salivation, respiratory difficulty, vomiting, and muscle twitching. (Usually, another anticholinesterase drug, pyridostigmine, is preferred for treating myasthenia gravis because it has fewer side effects.)

H₂-Receptor Antagonists

 $\rm H_2\text{-}receptor$ blockers such as ranitidine and nizatidine have prokinetic effects on intestinal smooth muscle in animals. These drugs are discussed later in Section Drugs for Treatment of Gastrointestinal Ulcers in Animals.

Erythromycin

Erythromycin is a macrolide antibiotic ordinarily used to treat bacterial infections. Pharmacology of macrolides is discussed in Chapter 36. It has long been associated with vomiting and regurgitation in small animals as an adverse consequence of treatment. This effect is caused by stomach contraction and expulsion at high doses. However, at low doses it can produce a beneficial stimulation of GI motility. Not all macrolide antibiotics exhibit this property because it requires a unique chemical structure that not all drugs in this class possess. (Erythromycin has a 14 carbon structure, but other macrolides that are less effective – tylosin and tilmicosin – have a 16 carbon structure.)

Erythromycin stimulates GI motility via activation of motilin receptors, via release of endogenous motilin, or via cholinergic mechanisms in the upper GI tract (Hall and Washabau, 1997; Lester et al., 1998; Hawkyard and Koerner, 2007). Motilin is a 22 amino acid peptide released from endocrine cells of duodenal mucosa. It increases the motor contractions, the housekeeper wave, during the interdigestive period. Motility is stimulated specifically in the pyloric antrum or the smooth muscle cells of the proximal small intestine (Nouri and Constable, 2007; Nouri et al., 2008). Because most of the motilin receptors are on the stomach and proximal small intestine, there is a weak response to erythromycin in the distal GI tract. In people, erythromycin has been used to promote gastric motility and increase stomach emptying in patients with diabetic gastroparesis and used in conjunction with enteral feeding in critical care patients (Hawkyard and Koerner, 2007).

The effective dose is 1 mg/kg or less – much lower than the antibacterial dose. It was effective for

stimulating motility in experimental horses (Ringger et al., 1996), but clinical responses to erythromycin in horses have been somewhat disappointing. One study showed that responses to erythromycin in horses that had undergone surgery were not as effective as the effects in healthy horses (Roussel et al., 2000). A dose of 8.8 mg/kg IM increased abomasal and rumen motility in calves (Nouri and Constable, 2007; Nouri et al., 2008; Wittek and Constable, 2005). The dose in small animals is also in the range of 0.5–1 mg/kg, but has not been tested for clinical efficacy (Whitehead et al., 2016). There is a concern that erythromycin may cause diarrhea in some horses through the effect on the normal bacterial flora of the intestine. An additional concern is that routine use may promote antibacterial resistance.

Lidocaine

Lidocaine is a well-known local anesthetic. (Local anesthetics are covered in more detail in Chapter 15, and with antiarrhythmics in Chapter 22.) It is used for local infiltration for minor surgical procedures and to treat cardiac arrhythmias. Intravenous infusions of lidocaine also improve intestinal motility in horses. Lidocaine has been used in horses postsurgically to reduce postoperative ileus. Postoperative ileus in horses is a widespread clinical problem that may be caused by (i) sympathetic stimulation, (ii) pain, or (iii) inflammation. These effects inhibit smooth muscle motility in the intestine and lidocaine may work by suppressing this transmission. Another view on the mechanism is that lidocaine does not have a direct prokinetic effect, but rather restores motility via other mechanisms (Cook and Bilkslager, 2008). These authors presented evidence that in horses lidocaine restores motility by inhibiting intestinal inflammation and reperfusion injury.

In one study (Malone et al., 2006), lidocaine administration to horses produced less reflux and shorter time of hospitalization. Infusions of lidocaine have decreased postoperative ileus either through a direct effect, or via suppression of painful stimuli. Doses in horses are 1.3 mg/kg loading dose (bolus), followed by 0.05 mg/kg/min IV infusion.

Adverse Effects

As with the other uses of lidocaine, systemic administration may produce adverse events. The most common in horses have been muscle fasciculations, ataxia, and seizures. If signs are observed, decrease rate of infusion.

Opiate Antagonists for Promoting Intestinal Motility

Opiates and their antagonists are discussed in Chapter 13. Activation of opiate μ receptors in the intestinal

1254 Veterinary Pharmacology and Therapeutics

smooth muscle decreases propulsive motility. Expression of μ -opiate receptors have been found in the submucosal plexus, myenteric plexus, and longitudinal muscle of the ileum. Activating these receptors has been used to treat some forms of diarrhea (e.g., loperamide). Administration of opiate analgesics postoperatively (Boscan et al., 2006; Sojka et al., 1988) or increased levels of endogenous opioids (endorphins), stimulate these receptors to inhibit intestinal motility causing postoperative ileus (DeHaven-Hudkins et al., 2008). Therefore, postoperative ileus may be treated by blocking intestinal opiate receptors (μ receptors) (Hicks et al., 2004).

Selective peripheral opiate antagonists act as peripheral opioid antagonists, rather than central opioid antagonists. They do not produce a central effect because they are unable to cross the blood-brain barrier. *Naloxone should not be used for this indication because it will cross the blood-brain barrier to diminish the analgesic effect of opioids.* Such agents include alvimopan, methylnaltrexone, and naloxegol.

Alvimopan (Entereg[®]) has advantages over methylnaltrexone with respect to potency and duration of activity (DeHaven-Hudkins et al., 2008; Taguchi et al., 2001). It is administered orally with low bioavailability (6%) and produces a local effect on the intestine to promote motility, without diminishing analgesic effect of opioids. It is a zwitterionic molecule and the high polarity restricts its diffusion across the blood–brain barrier. A dose of 3 mg orally to people, three times daily completely reversed the GI effects of morphine, without affecting analgesia. The typical dose is 12 mg (one capsule) administered orally prior to surgery, and continuing after surgery twice daily.

Methylnaltrexone (Relistor[®]) is available as a SC injection (0.15 mg/kg) administered once every 48 hours for postoperative ileus. Like alvimopan, it does not have systemic effects and will not interfere with analgesia. There has been limited use of methylnaltrexone in horses. At a dose of 0.75 mg/kg IV q 12 h for four days to horses inhibited morphine-induced intestinal effects (Boscan et al., 2006).

Naloxegol (Movantik[®]), in 12.5 and 25-mg tablets, is a pegylated opioid antagonist. It is used for oral treatment of opioid-induced constipation. It acts peripherally because pegylation of the molecule reduces the ability of naloxegol to cross the blood–brain barrier and makes it a substrate for the efflux transporter P-glycoprotein.

Drugs for Treatment of Gastrointestinal Ulcers in Animals

Histamine H_2 -receptor antagonists, sucralfate, proton pump inhibitors (omeprazole), and antacids remain

Table 46.3 Antiulcer drugs: clinical uses

Gastritis	
Gastric ulcers	
Duodenal ulcers	
Gastrointestinal ulcer prevention	
Esophagitis	
Mast cell tumors	
Hypergastrinemic syndromes	
Prevention and treatment of NSAID-induced ulcers	

the principal drugs used to manage gastrointestinal ulceration in small and large animals (Table 46.3; Figure 46.1). The medical management of ulcer diseases will not be covered in this section, but readers are referred to other references for this information (Merritt, 2003; Papich, 1993; Matz, 1995; Henderson and Webster, 2006a, 2006b; Feldman and Burton, 1990).

Because many of the ulcerative diseases encountered in veterinary medicine are induced by drugs that inhibit prostaglandin synthesis (nonsteroidal antiinflammatory drugs, NSAIDs), one should be familiar with the role of prostaglandins in the GI tract, how their synthesis is inhibited, and treatments used to maintain the protective effect of prostaglandins in the GI tract. Veterinarians also should be familiar with the normal physiological role of protective mucus layer in the stomach, the cytoprotective mechanisms, role of bicarbonate secretion, and the normal mechanisms that restore epithelial cells in the stomach and intestine. These factors were reviewed by Allen et al. (1993) several years ago, but are still relevant today. When these protective factors become disrupted or compromised, ulcers can occur in animals. Gastrointestinal ulcers are a major health problem in horses, pigs, dogs, cats, and zoo animals. Conditions that increase the risk of gastrointestinal ulceration are administration of ulcerogenic drugs (NSAIDs, corticosteroids, and stomach irritants), stress, disrupted mucosal blood supply, and inflammatory diseases.

Gastrointestinal ulceration is an important medical problem in horses, in which the prevalence in animals involved in showing and racing has been listed as 81– 93%, and even as high as 100% in some studies. In Thoroughbreds and Standardbreds the prevalence is was 80– 95%; and in show horses it may be as high as 58%. Factors such as stall confinement, intense exercise, diet (high energy concentration in diet), and racing stress may be contributing factors. Location of ulcers in horses is primarily in the squamous epithelium (nonglandular portion). Factors that contribute to ulcers are the intermittent feeding schedule and high stomach acidity. In sick foals, ulcers also are common. Factors that contribute to ulcers in foals are NSAIDs, stress, and sepsis.

1272 Veterinary Pharmacology and Therapeutics

Andersson T, Andren K, Cederberg C, Lagerstrom PO, Lundborg P, Skanberg I. (1990). Pharmacokinetics and bioavailability of omeprazole after single and repeated oral administration in healthy subjects. *Br J Clin Pharmacol.* **29**, 557–563.

Andersson T, Rohss K, Bredberg E, Hassan-Alin M. (2001). Pharmacokinetics and pharmacodynamics of esomeprazole, the S-isomer of omeprazole. *Aliment Pharmacol Therap.* **15**, 1563–1569.

Andrews FM, Doherty TJ, Blackford JT, Nadeau JA, Saxton AM. (1999). Effects of orally administered enteric-coated omeprazole on gastric acid secretion in horses. *Am J Vet Res.* **60**, 929–931.

Baker SJ, Gerring EL. (1993). Effects of single intravenously administered doses of omeprazole and ranitidine on intragastric pH and plasma gastrin concentration in nonfed ponies. *Am J Vet Res.* **54**, 2068–2074.

Barkun AN, Bardou M, Pham CQ, Martel M. (2012). Proton pump inhibitors vs. histamine 2 receptor antagonists for stress-related mucosal bleeding prophylaxis in critically ill patients: a meta-analysis. *Am J Gastroenterol.* **107**, 507–520.

Bartlett JG. (2002). Antibiotic-associated diarrhea. *N Engl J Med.* **346**, 334–339.

Bell NJ, Burget D, Howden CW, Wilkinson J, Hunt RH. (1992). Appropriate acid suppression for the management of gastro-oesophageal reflux disease. *Digestion.* 51 (Suppl. 1), 59–67.

Benchaoui HA, Cox SR, Schneider RP, Boucher JF, Clemence RG. (2007a). The pharmacokinetics of maropitant, a novel neurokinin type-1 receptor antagonist, in dogs. *J Vet Pharmacol Therap.* **30**, 336–344.

Benchaoui HA, Siedek EM, De La Puente-Redondo VA, Tilt N, Rowan TG, Clemence RG. (2007b). Efficacy of maropitant for preventing vomiting associated with motion sickness in dogs. *Vet Rec.* **161**, 444–447.

Bersenas AM, Mathews KA, Allen DG, Conlon PD. (2005). Effects of ranitidine, famotidine, pantoprazole, and omeprazole on intragastric pH in dogs. *Am J Vet Res.* **66**, 425–431.

Bierer DW. (1990). Bismuth subsalicylate: history, chemistry, and safety. *Rev Infect Dis.* **12** (Suppl.), S3–S8.

Boscan P, Monnet E, Mama K, Twedt DC, Congdon J, Steffey EP. (2011). Effect of maropitant, a neurokinin 1 receptor antagonist, on anesthetic requirements during noxious visceral stimulation of the ovary in dogs. *Am J Vet Res.* **72**, 1576–1579.

Boscan P, Van Hoogmoed LM, Farver TB, Snyder, Jr.
(2006). Evaluation of the effects of the opioid agonist morphine on gastrointestinal tract function in horses. *Am J Vet Res.* 67, 992–997.

Braun U, Steiner A, Kaegi B. (1990). Clinical, haematological and biochemical findings and the results of treat-ment in cattle with acute functional pyloric stenosis. *Vet Rec.* **126**, 107–110.

Burget DW, Chiverton SG, Hunt RH. (1990). Is there an optimal degree of acid suppression for healing of duodenal ulcers? A model of the relationship between ulcer healing and acid suppression. *Gastroenterology*. **99**, 345–351.

Castro JR, Adair HS, Radecki SV, Kiefer VR, Elliot SB, Longhofer SL. (2010). Effects of domperidone on digital laminar microvascular blood flow in clinically normal adult horses. *Am J Vet Res.* **71**, 281–287.

Chae JW, Song BJ, Baek IH, Yun HY, Ma JY, Kwon KI. (2015). Effects of food intake on pharmacokinetics of mosapride in beagle dogs. *J Vet Pharmacol Therap.* 38, 497–499.

Chiverton SG, Howden CW, Burget DW, Hunt RH. (1992). Omeprazole (20 mg) daily given in the morning or evening: a comparison of effects on gastric acidity, and plasma gastrin and omeprazole concentration. *Aliment Pharmacol Therap.* **6**, 103–111.

Claude AK, Dedeaux A, Chiavaccini L, Hinz S. (2014). Effects of maropitant citrate or acepromazine on the inci-dence of adverse events associated with hydromorphone premedication in dogs. *J Vet Intern Med.* **28**, 1414–1417.

Constable PD. (2004). Antimicrobial use in the treatment of calf diarrhea. *J Vet Intern Med.* **18**, 8–17.

Cook G, Papich MG, Roberts MC, Bowman KF. (1997). Pharmacokinetics of cisapride in horses after intrave-nous and rectal administration. *Am J Vet Res.* **58**, 1427–1430.

Cook VL, Blikslager AT. (2008). Use of systemically administered lidocaine in horses with gastrointestinal tract disease. *J Am Vet Med Assoc.* **232**, 1144–1148.

Court MH. (2013). Canine cytochrome P-450 pharmacogenetics. *Vet Clin North Am Small Anim Pract.* **43**, 1027–1038.

Dando TM, Perry CM. (2004) Aprepitant: A review of its use in the prevention of chemotherapy-induced nausea and vomiting. ADIS drug evaluation. *Drugs* **64**, 777–794.

Dandrieux JR, Noble PJM, Scase TJ, Cripps PJ, German AJ. (2013). Comparison of a chlorambucil-prednisolone combination with an azathioprine-prednisolone combination for treatment of chronic enteropathy with concurrent protein-losing enteropathy in dogs: 27 cases (2007–2010). J Am Vet Med Assoc. **242**, 1705–1714.

Danese S, Fiocchi C. (2011). Ulcerative colitis. *N Eng J Med.* **365**, 1713–1725.

Davis MS, Willard MD, Nelson SL, McCullough SM, Mandsager RE, Roberts J, Payton ME. (2003). Efficacy of omeprazole for the prevention of exercise-induced gastritis in racing Alaskan sled dogs. *J Vet Intern Med.* 17, 163–166. de Brito Galvao JF, Trepanier LA. (2008). Risk of hemolytic anemia with itravenous administration of famotidine to hospitalized cats. *J Vet Intern Med.* **22**, 325–329.

DeHaven-Hudkins DL, DeHaven RN, Little PJ, Techner LM. (2008). The involvement of the mu-opioid receptor in gastrointestinal pathophysiology: Therapeutic opportunities for antagonism at this receptor. *Pharmacol Ther.* **117**, 162–187.

de la Puente-Redondo VA, Tilt N, Rowan TG, Clemence RG. (2007a). Efficacy of maropitant for treatment and prevention of emesis caused by intravenous infusion of cisplatin in dogs. *Am J Vet Res.* **68**, 48–56.

de la Puente-Redondo VA, Siedek EM, Benchaoui HA, Tilt N, Rowan TG, Clemence RG. (2007b). The antiemetic efficacy of maropitant (Cerenia) in the treatment of ongoing emesis caused by a wide range of underlying clinical aetiologies in canine patients in Europe. *J Small Anim Pract.* **48**, 93–98.

de la Puente-Redondo V, Tingley FD 3rd, Schneider RP, Hickman MA. (2007c). The neurokinin-1 antagonist activity of maropitant, an antiemetic drug for dogs, in a gerbil model. *J Vet Pharmacol Therap.* **30**, 281–287.

Dillon R. (1989). Effects of glucocorticoids on the gastrointestinal system. In Kirk RW (ed.), *Current Veterinary Therapy X*. Philadelphia, WB Saunders Co. 897–904.

Egan LJ, Murray JA. (2000). New perspectives in gastric acid suppression: genetic polymorphisms predict the efficacy of proton pump inhibitors. *Dig Dis.* **18**, 58–63.

Feldman M, Burton ME. (1990). Histamine H2-receptor antagonists: Standard therapy for acid-peptic diseases. (Parts 1 and 2). N Engl J Med. 323, 1672–1680, 1749–1755.

Fukuda H, Koga T, Furukawa N, Nakamura E, Shiroshita Y. (1999). The tachykinin KN1 receptor antagonist
GR205171 abolishes the retching activity of neurons comprising the central pattern generator for vomiting in dogs. *Neurosci Res.* 33, 25–32.

Fukui H, Yamamoto M, Sato S. (1992). Vagal afferent fibres and peripheral 5-HT3 receptors mediate cispla-tin-induced emesis in dogs. *Jap J Pharmacol.* **59**, 221–226.

Furuta S, Kamada E, Suzuki T, Sugimoto T, Kawabata Y, Shinozaki Y, Sano H. (2001). Inhibition of drug metabolism in human liver microsomes by nizatidine, cimetidine and omeprazole. *Xenobiotica*. **31**, 1–10.

Gabardi S, Olyaei A. (2012). Evaluation of potential interactions between mycophenolic acid derivatives and proton pump inhibitors. *Ann Pharmacother*. **46**, 1054–1064.

Garcia-Mazcorro JF, Suchodolski JS, Jones KR, Clark-Price SC, Dowd SE, Minamoto Y, Markel M, Steiner JM, Dossin O. (2012). Effect of the proton pump inhibitor omeprazole on the gastrointestinal bacterial mi crobiota of healthy dogs. *FEMS Microbiol Ecol.* **80**, 624–636. Gerring EEL, Hunt JM. (1986). Pathophysiology of equine post-operative ileus: effects of adrenergic blockade, parasympathetic stimulation, and metoclopramide in an experimental model. *Equine Vet J.* **18**, 249–253.

Gerson LB, Triadafilopoulos G. (2001). Proton pump inhibitors and their drug interactions: an evidence-based approach. *Eur J Gastroenterol Hepatol.* **13**, 611–616.

Goetz TE, Oglivie GK, Keegan KG, Johnson PJ. (1990). Cimetidine for treatment of melanomas in three horses. *J Am Vet Med Assoc.* **196**, 449–452.

Gookin JL, Copple CN, Papich MG, Poore MF, Stauffer SH, Birkenheuer AJ, Twedt DC, Levy MG. (2006). Efficacy of ronidazole for treatment of feline *Tritrichomonas foetus* infection. *J Vet Intern Med.* **20**, 536–543.

Gould E, Clements C, Reed A, Giori L, Steiner JM, Lidbury JA, Suchodolski JS, Brand M, Moyers T, Emery L, Tolbert MK. (2016). A prospective, placebo-controlled pilot evaluation of the effect of omeprazole on serum calcium, magnesium, cobalamin, gastrin concentrations, and bone in cats. *J Vet Intern Med.* **30**, 779–786.

Grobman M, Reinero C. (2016). Investigation of neurokinin-1 receptor antagonism as a novel treatment for chronic bronchitis in dogs. *J Vet Intern Med.* **30**, 847–852.

Hall JA, Solie TN, Seim HB 3rd, Twedt DC. (1996). Effect of metoclopramide on fed-state gastric myoelectric and motor activity in dogs. *Am J Vet Res.* **57**, 1616–1622.

Hall JA, Washabau RJ. (1997). Gastrointestinal prokinetic therapy: motilin-like drugs. *Comp Contin Ed Pract Vet.* **19**, 281–288.

Hanauer SB. (1996). Inflammatory bowel disease. *N Engl J Med.* **334**, 841–848.

Hanson SM, Bostwick DR, Twedt DC, Smith MO. (1997). Clinical evaluation of cimetidine, sucralfate, and miso-prostol for prevention of gastrointestinal tract bleeding in dogs undergoing spinal surgery. *Am J Vet Res.* **58**, 1320–1323.

Haponen I, Linden J, Saari S, Karjalainen M, Hänninen ML, Jalava K, Westermarck E. (1998). Detection and effects of helicobacters in healthy dogs and dogs with signs of gastritis. *J Am Vet Med Assoc.* **213**, 1767–1774.

Hawkyard CV, Koerner RJ. (2007). The use of erythromycin as a gastrointestinal prokinetic agent in adult critical care: benefits versus risks. *J Antimicrob Chemother.* **59**, 347–358.

Hayashi A, Mizumoto T, Kusano T, Sekiguchi T, Itoh Z. (1990). Inhibition of gastric acid secretion by H-2 receptor antagonists normalizes interdigestive motor cycle in the stomach in dog and man. *Gastroenterology*. **98**, A56, 1990.

Hedner T, Cassuto J. (1987). Opioids and opioid receptors in peripheral tissue. *Scand J Gastrol.* **22** (Suppl.), 27–46.

Henderson AK, Webster CRL. (2006a). Disruption of the gastric mucosal barrier in dogs. *Comp Contin Ed Pract Vet.* **28**, 340–356.

Henderson AK, Webster CRL. (2006b). The use of gastroprotectants in treating gastric ulceration in dogs. *Comp Contin Ed Pract Vet.* 28, 358–372.

Hicks GA, DeHaven-Hudkins DL, Camilleri M. (2004). Opiates in the control of gastrointestinal tract function: current knowledge and new avenues for research. *Neurogastroenterol Motility*. **16** (Suppl. 2), 67–70.

Hickman MA, Cox SR, Mahabir S, Miskell C, Lin J, Bunger A, McCall RB. (2008). Safety, pharmacokinetics and use of the novel NK-1 receptor antagonist maropitant (CereniaTM) for the prevention of emesis and motion sickness in cats. *J Vet Pharmacol Therap.* **31**, 220–229.

Holland PS, Ruoff WW, Brumbaugh GW, Brown SA. (1997). Plasma pharmacokinetics of ranitidine HCL in adult horses. *J Vet Pharmacol Therap.* 20, 145–152.

Hostutler RA, Luria BJ, Johnson SE, Weisbrode SE, Sherding RG, Jaeger JQ, Guilford WG. (2004). Antibiotic-responsive histiocytic ulcerative colitis in 9 dogs. *J Vet Intern Med.* **18**, 499–504.

Huskey S-EW, Dean BJ, Doss GA, Wang Z, Hop CE, Anari R, Finke PE, Robichaud AJ, Zhang M, Wang B, Strauss JR. (2004). The metabolic disposition of aprepitant, a substance P receptor antagonist, in rats and dogs. *Drug Metab Dispos.* **32**, 246–258.

Hutchinson C, Geissler CA, Powell JJ, Bomford A. (2007). Proton pump inhibitors suppress absorption of dietary non-haem iron in hereditary haemochromatosis. *Gut.* **56**, 1291–1295.

Johnson SE. (1989). Loperamide: A novel antidiarrheal drug. *Comp Contin Ed.* **11**, 1373–1375.

Johnston KL, Lamport AI, Ballevre OP, Batt RM. (2000). Effects of oral administration of metronidazole on small intestinal bacteria and nutrients of cats. *Am J Vet Res.* **61**, 1106–1112.

Kempf J, Lewis F, Reusch CE, Kook PH. (2014). High-resolution manometric evaluation of the effects of cis-apride and metoclopramide hydrochloride administered orally on lower esophageal sphincter pressure in awake dogs. *Am J Vet Res.* **75**, 361–366.

Koh RB, Isaza N, Xie H, Cooke K, Robertson SA. (2014). Effects of maropitant, acepromazine, and electroacu-puncture on vomiting associated with administration of morphine in dogs. *J Am Vet Med Assoc.* **244**, 820–829.

Kraus BL. (2013). Efficacy of maropitant in preventing vomiting in dogs premedicated with hydromorphone. *Vet Anaesth Analges.* **40**, 28–34.

Kraus BL. (2014a). Efficacy of orally administered maropitant citrate in preventing vomiting associated with hydromorphone administration in dogs. *J Am Vet Med Assoc.* 244, 1164–1169.

Kraus BL. (2014b). Effect of dosing interval on efficacy of maropitant for prevention of hydromorphone-induced vomiting and signs of nausea in dogs. *J Am Vet Med Assoc.* 245, 1015–1020. Kromer W, Postius S, Riedel R. (2000). Animal pharmacology of reversible antagonism of the gastric acid pump, compared to standard antisecretory principles. *Pharmacology*. **60**, 179–187.

KuKanich K, KuKanich B. (2015). The effect of sucralfate tablets vs. suspension on oral doxycycline absorption in dogs. *J Vet Pharmacol Therap.* **38**, 169–173.

KuKanich K, KuKanich B, Guess S, Heinrich E. (2016). Effect of sucralfate on the relative bioavailability of enrofloxacin and ciprofloxacin in healthy fed dogs. *J Vet Intern Med.* **30**, 108–115.

Laine L, Hennekens C. (2010). Proton pump inhibitor and clopidogrel interaction: fact or fiction? *Am J Gastroenterol.* **105**, 34–41.

Larsson H, Carlsson E, Junggren U, Olbe L, Sjostrand SE, Skanberg NI, Sundell G. (1983). Inhibition of gastric acid secretion by omeprazole in the dog and rat. *Gastroenterology*. **85**, 900–907.

Lee RD, Vakily M, Mulford D, Wu J, Atkinson SN. (2009). Clinical trial: the effect and timing of food on the pharmacokinetics and pharmacodynamics of dexlansoprazole MR, a novel dual delayed release formulation of a proton pump inhibitor–evidence for dosing flexibility. *Aliment Pharmacol Therap.* **29**, 824–833.

LeGrange SN, Boothe DM, Herndon, Willard MD. (1997).
Pharmacokinetics and suggested oral dosing regimen of cisapride: a study in healthy cats. *J Am Anim Hosp Assoc.* 33, 517–523.

Leib MS, Hay WH, Roth L. (1989). Plasmacyticlymphocytic colitis in dogs. In Kirk RW. (ed.), *Current Veterinary Therapy X*. Philadelphia, WB Saunders Co. 939–944.

Lesman SP, Boucher JF, Grover GS, Cox SR, Bidgood TL. (2013). The pharmacokinetics of maropitant citrate dosed orally to dogs at 2 mg/kg and 8 mg/kg once daily for 14 days consecutive days. *J Vet Pharmacol Therap*. **36**, 462–470.

Lester GD, Merritt AM, Neuwirth L. (1998). Effect of erythromycin lactobionate on myoelectric activity of ileum, and cecal emptying of radiolabeled markers in clinically normal ponies. *Am J Vet Res.* **59**, 328–334.

Lester GD, Smith RL, Robertson ID. (2005). Effects of treatment with omeprazole or ranitidine on gastric squamous ulceration in racing Thoroughbreds. *J Am Vet Med Assoc.* **227**, 1636–1639.

Lindor K. (2007). Ursodeoxycholic acid for the treatment of biliary cirrhosis. *N Engl J Med.* **357**, 1524–1529.

Lo WK, Chan WW. (2013). Proton pump inhibitor use and the risk of small intestinal bacterial overgrowth: a meta-analysis. *Clin Gastroenterol Hepatol.* **31**, 483–490.

Lorenzutti AM, Martín-Flores M, Litterio NJ, Himelfarb MA, Zarazaga MP. (2016). Evaluation of the antiemetic efficacy of maropitant in dogs medicated with morphine and acepromazine. *Vet Anaesth Analg.* **43**, 195–198. Malone E, Ensink J, Turner T, Wilson J, Andrews F, Keegan K, Lumsden J. (2006). Intravenous continuous infusion of lidocaine for treatment of equine ileus. *Vet Surg.* **35**, 60–66.

Mansfield CS, James FE, Craven M, Davies DR, O'Hara AJ, Nicholls PK, Dogan B, MacDonough SP, Simpson KW. (2009). Remission of histiocytic ulcerative colitis in Boxer dogs correlates with eradication of invasive intramucosal Escherichia coli. *J Vet Intern Med.* **23**, 964–969.

Marlicz W, Łoniewski I, Grimes DS, Quigley EM. (2014). Nonsteroidal anti-inflammatory drugs, proton pump inhibitors, and gastrointestinal injury: contrasting interactions in the stomach and small intestine. *Mayo Clinic Proc.* **89**, 1699–1709.

Martin-Flores M, Sakai DM, Learn MM, Mastrocco A, Campoy L, Boesch JM, Gleed RD. (2016). Effects of maropitant in cats receiving dexmedetomidine and morphine. *J Am Vet Med Assoc.* **248**, 1257–1261.

Matz M. (1995). Antiulcer therapy. In Bonagura JD. (ed.), *Current Veterinary Therapy XII*. Philadelphia, WB Saunders Co. 706–710.

Mavligit GM. (1987). Immunologic effects of cimetidine: potential uses. *Pharmacotherapy*. 7 (Suppl.), 120S–124S.

Mealey KL. (2004). Therapeutic implications of the MDR-1 gene. *J Vet Pharmacol Therap.* **27**, 257–264.

Merritt AM. (2003). The equine stomach: a personal perspective. *AAEP Proc.* **49**, 75–102.

Mitchelson F. (1992). Pharmacological agents affecting emesis (Parts 1 and 2). *Drugs* **43**, 295–315, 443–463.

Morgan RV, Bachrach A. (1982). Keratoconjunctivitis sicca associated with sulfonamide therapy in dogs. *J Am Vet Med Assoc.* **180**, 432–434.

Mullin JM, Gabello M, Murray LJ, Farrell CP, Bellows J, Wolov KR, Kearney KR, Rudolph D, Thornton JJ. (2009).
Proton pump inhibitors: actions and reactions. *Drug Disc Today.* 14, 647–660.

Murray MJ. (1997). Suppression of gastric acidity in horses. *J Am Vet Med Assoc.* **211**, 37–40.

Narishetty ST, Galvan B, Coscarelli E, Aleo M, Fleck T, Humphrey W, McCall RB. (2009). Effect of refrigeration of the antiemetic Cerenia (maropitant) on pain on injection. *Vet Therap.* **10**, 93–102.

Neiger R, Simpson KW. (2000). Helicobacter infection in dogs and cats: facts and fiction. *J Vet Intern Med.* **14**, 125–133.

Nieto JE, Spier S, Pipers FS, Stanley S, Aleman MR, Smith DC, Snyder JR. (2002). Comparison of paste and suspension formulations of omeprazole in the healing of gastric ulcers in racehorses in active training. *J Am Vet Med Assoc.* **221**, 1139–1143.

Nieto JE, Maher O, Stanley SD, Larson R, Snyder JR. (2013). In vivo and in vitro evaluation of the effects of

domperidone on the gastrointestinal tract of healthy horses. *Am J Vet Res.* **74**, 1103–1110.

- Nouri M, Constable PD. (2007). Effect of parenteral administration of erythromycin, tilmicosin, and tylosin on abomasal emptying rate in suckling calves. *Am J Vet Res.* **68**, 1392–1398.
- Nouri M, Hajikolaee MR, Constable PD, Omidi A. (2008). Effect of erythromycin and gentamicin on abomasal emptying rate in suckling calves. *J Vet Intern Med.* **22**, 196–201.
- Papich MG. (1993). Antiulcer therapy. *Vet Clin North Am Small Anim Prac.* 23, 497–512.

Papich MG, Davis CA, Davis LE. (1987). Absorption of salicylate from an antidiarrheal preparation in dogs and cats. *J Am Anim Hosp Assoc.* **23**, 221–226.

Parkinson S, Tolbert K, Messenger K, Odunayo A, Brand M, Davidson G, Peters E, Reed A, Papich MG. (2015).
Evaluation of the effect of orally administered acid suppressants on intragastric pH in cats. *J Vet Intern Med.* 29, 104–112.

Prichard PJ, Yeomans ND, Mihaly GW, Jones BD, Buckle PJ, Smallwood RA, Louis WJ. (1985). Omeprazole: a study of its inhibition of gastric pH and oral pharmacokinetics after morning or evening dosage. *Gastroenterology*. **88**, 64–69.

Quimby JM, Brock WT, Moses K, Bolotin D, Patricelli K. (2015). Chronic use of maropitant for the management of vomiting and inappetence in cats with chronic kidney disease: a blinded placebo-controlled clinical trial. *J Feline Med Surg.* **17**, 692–697.

Quimby JM, Gustafson DL, Lunn KF. (2011). The pharmacokinetics of mirtazapine in cats with chronic kidney disease and in age-matched control cats. *J Vet Intern Med.* **25**, 985–989.

Quimby JM, Lunn KF. (2013). Mirtazapine as an appetite stimulant and anti-emetic in cats with chronic kidney disease: A masked placebo-controlled crossover clinical trial. *Vet J.* **197**, 651–655.

Ringger NC, Lester GD, Neuwirth L, Merritt AM, Vetro T, Harrison J. (1996). Effect of bethanechol or erythromycin on gastric emptying in horses. *Am J Vet Res.* **57**, 1771–1775.

Roger T, Bardon T, Ruckebusch Y. (1985). Colonic motor responses in the pony: Relevance of colonic stimulation by opiate antagonists. *Am J Vet Res.* **46**, 31–35.

Roussel AJ, Hooper RN, Cohen ND, Bye AD, Hicks RJ, Bohl TW. (2000). Prokinetic effects of erythromycin on the ileum, cecum, and pelvic flexure of horses during the postoperative period. *Am J Vet Res.* **61**, 420–424.

Sams RA, Gerken DF, Dyke TM, Reed SM, Ashcraft SM. (1997). Pharmacokinetics of intravenous and intra-gastric cimetidine in horses. 1. Effects of intravenous cimetidine on pharmacokinetics of intravenous phenylbutazone. *J Vet Pharmacol Therap.* 20, 355–361.

Sanchez LC, Lester GD, Merritt AM. (1998). Effect of ranitidine on intragastric pH in clinically normal neonatal foals. *Am J Vet Res.* **212**, 1407–1412.

Sasaki N, Okamura K, Yamada H. (2005). Effects of mosapride, a 5-hydroxytryptamine 4 receptor agonist, on electrical activity of the small intestine and cecum in horses. *Am J Vet Res.* **66**, 1321–1323.

Simpson KW, Neiger R, DeNovo R, Scherding. (2000). The relationship of Helicobacter spp. infection to gastric disease in dogs and cats. *J Vet Intern Med.* **14**, 223–227.

Sojka JE, Adams SB, Lamar CH, Eller LL. (1988). Effect of butorphanol, pentazocine, meperidine, or metoclopramide on intestinal motility in female ponies. *Am J Vet Res.* **49**, 527–529.

Solcia EN. (1993). Long-term omeprazole therapy in peptic ulcer disease: gastrin, endocrine cell growth, and gastritis. *Gastroenterology*. **104**, 1356–1370.

Suerbaum S, Michetti P. (2002). Heliobacter pylori infection. *N Engl J Med.* **347**, 1175–1186.

Sutalo S, Ruetten M, Hartnack S, Reusch CE, Kook PH. (2015). The effect of orally administered ranitidine and once-daily or twice-daily orally administered omeprazole on intragastric pH in cats. *J Vet Intern Med.* 29, 840–846.

Sykes BW, Sykes KM, Hallowell GD. (2014). A comparison of two doses of omeprazole in the treatment of equine gastric ulcer syndrome: A blinded, randomised, clinical trial. *Equine Vet J.* **46**, 416–421.

Sykes BW, Sykes KM, Hallowell GD. (2015a). A comparison of three doses of omeprazole in the treatment of equine gastric ulcer syndrome: A blinded, randomised, dose–response clinical trial. *Equine Vet J.* **47**, 285–290.

Sykes BW, Underwood C, Greer R, McGowan CM, Mills PC. (2016). Pharmacokinetics and bioequivalence testing of five commercial formulations of omeprazole in the horse. *J Vet Pharmacol Therap.* **39**, 78–83.

Sykes BW, Underwood C, McGowan CM, Mills PC. (2015b). Pharmacokinetics of intravenous, plain oral and enteric-coated oral omeprazole in the horse. *J Vet Pharmacol Therap.* **38**, 130–136.

Taguchi A, Sharma N, Saleem RM, Sessler DI, Carpenter RL, Seyedsadr M, Kurz A. (2001). Selective postoperative inhibition of gastrointestinal opioid receptors. *N Engl J Med.* **345**, 935–940.

Thames BE, Lovvorn J, Papich MG, Wills R, Archer T, Mackin A, Thomason J. (2017). The effects of clopidogrel and omeprazole on platelet function in normal dogs. *J Vet Pharmacol Ther.* **40**, 130–139.

Tolbert K, Bissett S, King A, Davidson G, Papich M, Peters E, Degernes L. (2011). Efficacy of oral famotidine and 2 omeprazole formulations for the control of intragastric pH in dogs. *J Vet Intern Med.* **25**, 47–54.

Tolbert K, Odunayo A, Howell R, Peters EE, Reed A. (2015). Efficacy of intravenous administration of

combined acid suppressants in healthy dogs. *J Vet Intern Med.* **29**, 556–560.

Tumulty JW, Broussard JD, Steiner JM, Peterson ME, Williams DA. (2004). Clinical effects of short-term oral budesonide on the hypothalamic-pituitary-adrenal axis in dogs with inflammatory bowel disease. *J Am Anim Hosp Assoc.* 40, 120–123.

Vail DM, Rodabaugh HS, Conder GA, Boucher JF, Mathur S. (2007). Efficacy of injectable maropitant (Cerenia) in a ramdomized clinical trial for prevention and treatment of cisplatin-induced emesis in dogs presented as veterinary patients. *Vet Comp Oncol.* **5**, 38–46.

Van Nueten JM, Schuurkes JAJ. (1992). Development of a gastrointestinal prokinetic: pharmacology of cisapride. *Front Gastroenterol Res.* **20**, 54–63.

Vemuri VK, Makriyannis A. (2015). Medicinal chemistry of cannabinoids. *Clin Pharmacol Therap.* 97, 553–558.

Wallace JL, Syer S, Denou E, de Palma G, Vong L, McKnight W, Jury J, Bolla M, Bercik P, Collins SM, Verdu E. (2011). Proton pump inhibitors exacerbate NSAID-induced small intestinal injury by inducing dysbiosis. *Gastroenterology*. **141**, 1314–1322.

Wallmark B, Larsson H, Humble L. (1985). The relationship between gastric acid secretion and gastric H+,K+-ATPase activity. *J Biol Chem.* **260**, 13681–13684.

- Washabau RJ, Hall JA. (1995). Cisapride. J Am Vet Med Assoc. 207, 1285–1288.
- Washabau RJ, Hall JA. (1997). Diagnosis and management of gastrointestinal motility disorders in dogs and cats. *Comp Contin Ed Pract Vet.* **19**, 721–736.

Washabau RJ, Holt DE. (2005). Diseases of the large intestine. In Ettinger SJ, Feldman ED. (eds), *Textbook of Veterinary Internal Medicine*, 6th edn. St Louis, Elsevier-Saunders, 1378–1407.

Washabau RJ, Summarco J. (1996). Effects of cisapride on feline colonic smooth muscle function. *Am J Vet Res.* **57**, 541–546.

Watson JW, Gonsalves SF, Fossa AA, McLean, Obach S, Andrews PL. (1995). The anti-emetic effects of CP-99,994 in the ferret and the dog: role of the NK1 receptor. *Br J Pharmacol.* **115**, 84–94.

Westermarck E, Frias R, Skrzypczak T. (2005a). Effect of diet and tylosin on chronic diarrhea in beagles. *J Vet Intern Med.* **19**, 822–827.

Westermarck E, Skrzypczak T, Harmoinen J, Steiner JM, Ruaux CG, Williams DA, Eerola E, Sundback P, Rinkinen M. (2005b). Tylosin-responsive chronic diarrhea in dogs. *J Vet Intern Med.* **19**, 177–186.

Whitehead K, Cortes Y, Eirmann L. (2016). Gastrointestinal dysmotility disorders in critically ill dogs and cats. *J Vet Emerg Crit Care*. **26**, 234–253.

Wilson DV, Evans AT, Mauer WA. (2006). Influence of metoclopramide on gastroesophageal reflux in anesthetized dogs. *Am J Vet Res.* 67, 26–31.

- Winberg B, Spohr A, Dietz HH, Egelund T, Greiter-Wilke A, McDonough SP, Olsen J, Priestnall S, Chang YF, Simpson KW. (2005). Quantitative analysis of inflammatory and immune response in dogs with gastritis and their relationship to Helicobacter spp. infection. *J Vet Intern Med.* **19**, 4–14.
- Wittek T, Constable PD. (2005). Assessment of the effects of erythromycin, neostigmine, and metoclo-pramide on abomasal motility and emptying rate in calves. *Am J Vet Res.* **66**, 545–552.
- Wu Y, Loper A, Landis E, Hettrick I, Novak L, Lynn K, Chen C, Thompson K, Higgins R, Batra U, Shelukar S, Kwei G, Storey D. (2004). The role of biopharmaceutics in the development of a clinical nanoparticle formulation of MK 0869: a Beagle dog model predicts improved bioavailability and diminished food effect on absorption in human. *Int J Pharm.* **285**, 135–146.
- Yamasaki K, Suematsu H, Takahashi T. (1998). Comparison of gastric lesions in dogs and cats with and without gastric spiral organisms. *J Am Vet Med Assoc.* **212**, 529–533.
- Yasuda S, Horai Y, Tomono Y, Nakai H, Yamato C, Manabe K, Kobayashi K, Chiba K, Ishizaki T. (1995). Comparison of the kinetic disposition and metabolism of E3810, a new proton pump inhibitor, and omeprazole in relation to S-mephenytoin 4'-hydroxylation status. *Clin Pharmacol Therap.* **58**, 143–154.
- Zhou R, Moench P, Heran C, Lu X, Mathias N, Faria TN, Wall DA, Hussain MA, Smith RL, Sun D. (2005).
 pH-dependent dissolution in vitro and absorption in vivo of weakly basic drugs: development of a canine model. *Pharm Res.* 22, 188–192.